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Regioselectivity in Alkylation Reactions of 1,2-O-Stannylene Acetals of D-Arabinofuranose

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ABSTRACT

The synthesis of β -arabinofuranosides via alkylation of 1,2-*O*-stannylene acetal intermediates has been studied. With reactive alkyl halides (benzyl bromide, allyl bromide, and *p*-methoxybenzyl chloride), the method provides a mixture of β -arabinofuranosides and 2-*O*-alkylated lactols in ratios of 4:1 to 1:1.5. However, with carbohydrate-derived electrophiles, no alkylated products are produced. It appears, therefore, that the method is limited to the preparation of β -arabinofuranosides of simple alcohols. Through the use of computational chemistry, we have explored the conformational properties of one of these stannylene acetals and propose that these species exist in more than one conformation in solution and that this contributes to the relatively poor regioselectivity in these reactions.

Key Words: Stannylene acetal; β-arabinofuranosides; Glycosylation; Alkylation.

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Scheme 1. Representative examples of a β -arabinofuranoside (1), a β -fructofuranoside (2) and a β -mannopyranoside (3).

INTRODUCTION

Polysaccharides containing β -arabinofuranosyl residues (**1**, Scheme 1) are important structural components of the mycobacterial cell wall.^[1,2] Recent interest^[3,4] in identifying inhibitors of the enzymes involved in the biosynthesis of these glycans has prompted the development of new methods for the synthesis of β -arabinofuranosides,^[5-10] as convenient access to fragments of these polysaccharides is needed for fundamental biochemical investigations aimed ultimately at drug and vaccine development.^[11-15] The stereoselective synthesis of β -arabinofuranosides, as well as the related β -fructofuranosides (**2**), is plagued by the same problems that complicate the preparation of the stereochemically analogous β -mannopyranosides (**3**).^[16,17] However, in contrast to the significant body of work that has addressed the stereoselective synthesis of β -mannopyranosides,^[16,17] the preparation of 1,2-*cis*- β -furanosides has received relatively little attention until recently.^[5-10,18-20]

Shown in Figure 1 is a straightforward approach for the synthesis of β -mannopyranosides (or β -rhamnopyranosides) that was developed by Hodosi and Kovac.^[21,22] The method involves the treatment of a reducing sugar (4) with dibutyltin oxide to produce the corresponding stannylene acetal (5), which is subsequently reacted with an electrophile to generate the β -glycoside (6) in a stereoselective fashion. This work extends earlier investigations on the alkylation of 1,2-*O*-stannylene acetals with simple alkyl halides, which led to the production of β -mannopyranosides,^[23,24] α -glucopyranosides,^[23] and α -ribofuranosides.^[25] The simplicity of this approach, coupled with the ready availability of 3,5-protected arabinofuranose derivatives (e.g., 7, Figure 2), prompted us to explore this method for the synthesis of β -arabinofuranosides.



Figure 1. Stannylene acetal approach to β -mannopyranosides developed by Hodosi and Kovác.

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Figure 2. Proposed synthesis of β -arabinofuranosides via the stannylene acetal approach.

RESULTS AND DISCUSSION

In exploring the potential of the route shown in Figure 2 for the synthesis of β -arabinofuranosides, we chose to probe the effect that the O-3 and O-5 protecting groups had on the outcome of the reaction. We therefore chose diols **10–12** (Scheme 2) as substrates. Compounds **10**^[26] and **11**^[27] were synthesized as previously reported. The preparation of **12** (Figure 3) was achieved from the known thioglycoside **13**,^[28] upon treatment with *N*-iodosuccinimide and silver triflate in wet acetonitrile. This reaction afforded **12** in 78% yield as a 1:2.3 α : β mixture of isomers.

With all three substrates in hand, we explored the synthesis of simple alkyl glycosides via the route outlined in Figure 2. A number of reaction conditions were explored, and we found that the best results were obtained when the glycosylation was carried out by heating a solution of the diol (e.g., **10**) with dibutyltin oxide in toluene at reflux for 30 minutes, followed by addition of the electrophile and n-Bu₄NBr. The progress of the reaction was monitored by TLC and was stopped after no change was observed. In our initial series of reactions, we employed *p*-methoxybenzyl chloride, benzyl bromide, or allyl bromide as the electrophile. The results of these reactions are presented in Table 1; the structures of the products are provided in Scheme 3.

From the data presented in Table 1 it is clear that this approach is of limited utility for the stereocontrolled synthesis of β -arabinofuranosides in high yield. As hoped, the reaction produces the desired β -glycoside (e.g., **14**) and none of the corresponding α -glycoside was detected. However, in all cases, the reaction also produces significant quantities of the isomeric 2-*O*-alkylated product (e.g., **15**) as a byproduct. These lactol byproducts were each isolated as an α/β mixture that was inseparable by chromatography.



Scheme 2. Glycosyl donors 10–12.



Figure 3. Synthesis of 12, (a) NIS, AgOTf, CH₃CN, H₂O, 0 °C, 78%.

¹H and ¹³C NMR spectroscopy were used to determine the structures of the products. In the ¹H NMR spectra of the β -glycosides, the resonance for the anomeric hydrogen appeared as a doublet ($J \sim 4.5$ Hz) between 5.0 and 5.2 ppm. The ¹³C NMR spectra of these compounds showed a resonance for the anomeric carbon in the range of 99–102 ppm. Both of these features are diagnostic for β -arabinofuranosides.^[29] Had the α -glycosides been produced, the resonances for the anomeric hydrogens would have appeared in the ¹H NMR spectra as singlets, or doublets with a smaller coupling constant ($J \sim 1.5$ Hz) and the resonances for the anomeric carbons would have been found in the ¹³C NMR spectra above 105 ppm.^[29] The identity of the lactols was easily ascertained from ¹³C NMR spectra, which showed two resonances in the region

Table 1. Use of 1,2-O-stannylene acetals in the synthesis of β -arabinofuranosides.

$\begin{array}{c} \text{PO} & \text{OH} \\ \text{O} & \text{OH} \\ \text{O} & \text{OH} \end{array} \\ \begin{array}{c} \text{Bu}_2 \text{SnO} \\ \text{toluene, reflux} \end{array}$	n-Bu-Sn PO-O- PO-O- PO-O-	RX <i>n</i> -Bu ₄ NBr PO PO +	PO O PO PO PO
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Substrate	Alkyl	Reaction	Isolated	Products (ratio) ^a
	halide	time (h)	yield (%)	
10	AllBr	3	52	14:15 (1.7:1)
10	BnBr	1	61	16:17 (1.8:1)
10	PMBCl	0.25	72	18:19 (1:1)
11	AllBr	3	20	20:21 (2:1)
11	BnBr	3	42	22:23 (4:1)
11	PMBCl	3	59	24:25 (1.5:1)
12	AllBr	7	59	26:27 (1:1.5)
12	BnBr	7	73	28:29 (1.3:1)
12	PMBCl	7	81	30:31 (1.3:1)

^aRatio determined from mass of isolated products.

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Scheme 3. Structures of products produced in alkylation reactions.

between 94 and 102 ppm, corresponding to C-1 in the α and β reducing sugars. The presence of the two isomers was also clearly visible in the ¹H NMR spectrum; however, due to spectral overlap it was sometimes difficult to assign the signals arising from the anomeric hydrogen in both anomers. In the case of **17**, further proof for the structure was obtained by comparison with the ¹H and ¹³C NMR data for the known compound,^[30] which is commercially available.

The overall alkylation yields ranged from 20-80% and the best results were obtained when *p*-methoxybenzyl chloride was used as the electrophile. With regard to the donor substrates, the benzylated and silylated donors **10** and **12** gave modest to good combined yields of the alkylated products. The yields were slightly better with **12** as compared to **10**; however, the regioselectivity in favor of the β -glycoside was marginally higher when **10** was used as the donor. It is perhaps not unexpected that the benzoylated substrate, **11**, gave the lowest overall yields. The electron-withdrawing ability of the ester protecting groups on **11** would be expected to reduce the nucleophilicity of the stannylene acetal oxygens through an inductive effect.

To rationalize the regioselectivity of these reactions, we considered first the factors that govern alkylations of stannylene acetals. In reactions involving stannylene acetals formed from non-anomeric *cis*-diols on pyranose rings, the equatorial oxygen is almost always the site of predominant reaction.^[31,32] Although these stannylene acetals are known to aggregate in solution, when halide salts are added to promote the reaction (as in the present case), it is believed that they react through their monomeric forms.^[31] An important consideration is that the relative rigidity of the pyranose ring ensures that the molecule adopts a single major conformation in solution, at least with respect to the relative orientation of O-1 and O-2, and thus the regioselectivity is generally high. However, anomalous results have been obtained. For example, Srivastava and Schuerch reported^[23] that reaction of methyl iodide with stannylene acetal **32** (Scheme 4) provided a 7:3 mixture of α -glycoside **33** and the 2-*O*-alkylated lactol **34**. It was subsequently shown^[22] that treatment of triflate **35** with the maltose-derived stannylene



Scheme 4. Compounds 32-37.

acetal **36** yielded the 2-*O* alkylated product **37** in 75% yield and no mention was made of the formation of the corresponding glycoside.

The paucity of previous investigations on organotin-mediated reactions on furanose rings makes it difficult to draw substantially on literature precedence when proposing an explanation for the regioselectivity observed in the alkylation reactions of these arabinofuranose systems. Although some of the first demonstrated examples of stannylene acetals in protecting group chemistry were carried out on nucleosides,^[33] relatively little additional work has been done on these systems to date.^[25,34,35] In the initial paper by Moffat and coworkers,^[33] acylations of ribonucleosides (e.g., **38**, Scheme 5) via stanylene acetals were investigated. In general, alkylation of O-3 predominated (e.g., **39**), however, determining the inherent regioselectivity of the process was complicated by the tendency of the acyl group on O-2 to migrate to O-3. A paper more directly relevant to the present work describes the alkylation of stannylene acetal **40**.^[36,37] When methyl iodide was used as the electrophile, a mixture of 2-*O*-methyl lactol (**41**) and α -glycoside (**42**) were produced in 1:1 ratio, whereas benzyl bromide afforded the α -glycoside as the major product over the corresponding lactol (83%:13%).

Furanose rings are inherently more flexible than their pyranose counterparts, and the substituents on a five membered ring adopt pseudo-axial or pseudo-equatorial orientations rather than classical axial or equatorial arrangements. Mindful of this issue,



Scheme 5. Compounds 38-42.



Scheme 6. Comparison of 43 and 44.

we envisioned two possible explanations for the poor regioselectivity observed in these alkylation reactions. One possibility is that in the stannylene acetal intermediate the pseudo-axial/pseudo-equatorial placement of both oxygens does not lead to sufficient steric differentiation and therefore they compete for the electrophile. Another plausible explanation is that the stannylene acetal adopts two or more low energy conformations in which the relative orientations of O-1 and O-2 differ, e.g., O-1 is pseudo-axial in one conformer and pseudo-equatorial in another. An erosion in regioselectivity would be expected to result from the reaction of both conformers with the electrophile. Under either scenario, it could be expected that changes in reaction conditions (e.g., protecting groups on the donor, the electrophile used) would influence the regioselectivity of the reaction, as is observed in the results presented in Table 1.

In an effort to probe these hypotheses we carried out density functional theory^[36,37] calculations on stannylene acetal, **43** (Scheme 6), the species formed upon reaction of **12** with dibutyltin oxide. This substrate was chosen as the yields of these reactions were the best of the three donors explored. The general flexibility of the ring was of concern and so we began by generating a diverse family of conformers using the Systematic Pseudo Monte Carlo (SPMC) search protocol as implemented in Macro-Model version 6.5.^[38,39] A limitation in the application of MacroModel to this problem is that it is not parameterized for tin and therefore the SPMC search was carried out on an analog (**44**, Scheme 6) in which the tin atom was replaced with silicon. As the purpose of the search was simply to generate diverse geometries for higher level calculations, we viewed the tin/silicon replacement as a reasonable alternative to the manual generation of a large number of initial starting geometries via an approach that would necessarily be biased by intuition.

A total of 10,000 structures of **44** were generated by the SPMC search, which were then minimized in the gas phase at the AM1^[40] level of theory, yielding a family of 113 conformers, all of which were within 32 kcal/mol of the lowest energy structure. Clustered around the global minimum was a group of four structures that all possessed similar furanose ring conformations. A second family of six conformers, all possessing a common furanose ring conformation, was present ~ 1.1 kcal/mol above the global minimum (see Experimental Section for additional details). The other 103 conformers obtained after AM1 optimization were more than 3.0 kcal/mol above the global minimum structure. The lowest AM1 energy structures from each of these two families of conformers were then taken, the silicon atom connecting O-1 and O-2 replaced with tin, and the resulting structures were subjected to higher level calculations as outlined in the Experimental section. The optimized geometries for these conformers (**43-A** and **43-B**) are shown in Scheme 7 (all hydrogens and the alkyl groups on the siloxane moiety are omitted for clarity). The furanose ring in **43-A** adopts an ${}^{O}E$ conformation, which places O-1 pseudo-equatorial and O-2 pseudo-axial; in **43-B** the ring is in a ${}^{3}T_{2}$ conformation, in which O-2 is pseudo-equatorial and O-1 is pseudo-axial. Thus, the orientation of O-1 and O-2 differ in these two structures. Based upon our calculations, these two conformers differ by 2.7 kcal/mol, with **43-A** being of lower energy.

These calculations suggest that the furanose ring in stannylene acetal **43** exists in (at least) two conformations in which the relative pseudo-axial/pseudo-equatorial orientations of O-1 and O-2 differ. Moreover, the differences in energy between these conformers, **43-A** and **43-B**, are relatively small and, given the relatively low interconversion barriers typically seen for furanose rings,^[41] it is reasonable to expect that these isomers interconvert rapidly, especially under the elevated temperatures at which these reactions are carried out. Taken together, these data suggest that the low regioselectivity observed in the alkylation reactions in these systems is the result of the presence of more than one stannylene acetal conformer, which react with electrophiles at O-1 and O-2 to varying degrees. We note that we have examined only the thermodynamically most stable conformers and it is possible (likely) that other reactive conformers may also be present. Under these conditions, it is plausible that the regioselectivity of the reaction is influenced not only by these conformational effects, but also by the donor substrates and reaction conditions. For example, in the results presented in Table 1, the benzoylated



Scheme 7. B3LYP-optimized geometries of **43-A** and **43-B**. For clarity, all hydrogen atoms as well as carbon atoms of the siloxane acetal and butyl groups have been deleted.

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Scheme 8. Compounds 45-49.

donor **11** gave the highest regioselectivity in the alkylation reactions. The electronwithdrawing ability of the benzoyloxy group at C-3 can be invoked to rationalize this observation. The closer proximity of this protecting group to O-2 would be expected to decrease the nucleophilicity of this oxygen relative to O-1.

Despite the relatively modest results in the alkylation reactions with the simple electrophiles, we explored the possibility of synthesizing oligosaccharides via this approach. As a simple model carbohydrate electrophile, we chose triflate 45 (Scheme 8), which was prepared from the known alcohol $46^{[42]}$ upon treatment with triflic anhydride and pyridine. However, upon reaction of 45 with the stannylidene acetal generated from 12, no alkylated products could be isolated. Earlier reports^[22] have demonstrated that polar solvents can also be used to promote these alkylation reactions. The reaction was therefore repeated by generating the stannylene acetal and then adding 45 and DMF. However, when these conditions were used, no disaccharide was formed; rather, the formate ester 47 was produced in modest (39%) yield. The production of formate esters as byproducts in these reactions has been previously reported^[22] and has been minimized by carrying out the reaction at reduced temperatures. However, in our hands, cooling the reaction mixture to room temperature led to the formation of no alkylated products. As a final attempt to prepare a disaccharide, we synthesized the iodide 49 (via tosylate 48) and reacted it with stannylene acetal 12 and n-Bu₄NBr in toluene at reflux. However, no alkylated products were produced.

CONCLUSIONS

In conclusion, the stereoselective synthesis of β -arabinofuranosides in high yields was shown to be unsuccessful via of the 1,2-*O*-stannylene approach. Although the formation of the α -glycoside was avoided, our results with simple electrophiles demonstrated that the reactivity difference between the O-1 and O-2 is small. We propose that in these furanose ring systems, more than one reactive stannylene acetal conformer is present and they collectively contribute to the decrease in regioselectivity observed in these alkylation reactions. This contrasts to the application of this method to the synthesis of β -mannopyranoside or β -rhamnopyranosides, where the rigid nature of the pyranose ring orients the tin acetal oxygen atoms in distinctly different steric environments. In the course of our investigations, Oscarsson and coworkers reported a study in which the stereoselective synthesis of β -fructofuranosides (e.g., **2**, Scheme 1) via stannylene acetals was explored.^[18] When a benzylated species analogous to **10** was used as the donor and allyl bromide was employed as the electrophile, the desired fructofuranosides were produced as a 1:4.6 α/β mixture. Interestingly, no products arising from alkylation at the non-anomeric oxygen were reported; however, as in our investigations, the use of carbohydrate electrophiles gave no disaccharide products. These results, together with ours and earlier work by Klein^[25] and coworkers, suggest the 1,2-*O*-stannylene acetal glycosylation methodology is of limited utility for the stereocontrolled synthesis of 1,2-*cis*-furanosides in high yield.

EXPERIMENTAL

Solvents were distilled from the appropriate drying agents before use. Unless stated otherwise, all reactions were carried out under a positive pressure of argon and were monitored by TLC on silica gel 60 F_{254} (0.25 mm, E. Merck). Spots were detected under UV light, by charring with 10% H_2SO_4 in ethanol, or by charring with anisaldehyde in ethanol. Solvents were evaporated under reduced pressure and below 40°C (bath). Column chromatography was performed on silica gel 60 (40–60 μ M). The ratio between silica gel and crude product ranged from 100 to 50:1 (w/w). Optical rotations were measured at 21 ± 2°C. ¹H NMR spectra were recorded at 400 or 500 MHz, and chemical shifts are referenced to TMS (0.0, CDCl₃). ¹³C NMR spectra were recorded at 100 or 125 MHz, and ¹³C chemical shifts are referenced to CDCl₃ (77.00, CDCl₃). Elemental analyses were performed by Atlantic Microlab Inc., Norcross, GA. Electrospray mass spectra were recorded on samples suspended in THF or CH₃OH with added NaCl.

3,5-*O*-(**Tetraisopropylsiloxane-1,3-diyl**)-**D**-**arabinofuranose** (12). Thioglycoside **13** (990 mg, 2.0 mmol) was dissolved in acetonitrile (10 mL) and water (2 mL) and the mixture was cooled to 0°C. *N*-iodosuccinimide (0.49 g, 2.2 mmol) and silver triflate (0.13 g, 0.5 mmol) were added to the reaction mixture and, after stirring for 5 min, triethylamine (1.0 mL) was added and the mixture was filtered through Celite. The filtrate was diluted with CH₂Cl₂, washed with water, and the organic layer was dried (MgSO₄) and concentrated. The residue was purified by chromatography (hexane/ EtOAc, 2:1) to give **12** (600 mg, 78%) as a clear syrup: R_f 0.3 (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, $\delta_{\rm H}$) 5.25 (d, 0.7 H, *J* = 4.7 Hz), 5.22 (d, 0.3 H, *J* = 2.8 Hz), 4.31–3.63 (m, 6 H), 2.70 (br s, 1 H), 1.10–0.94 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, $\delta_{\rm C}$) 101.9, 95.0, 82.8, 81.7, 78.6, 78.4, 78.1, 73.0, 66.3, 65.3, 17.9 (2), 17.80 (2), 17.7 (3), 17.5 (2), 17.4 (4), 13.9, 13.6, 13.2, 12.9 (2).

Anal. Calcd for $C_{17}H_{36}O_6Si_2$ (398.1366): C, 52.00; H, 9.24. Found: C, 51.98; H, 9.21.

General procedure for glycosylation reactions. A mixture of the diol (1.3 mmol), and dibutyltin oxide (1.5 mmol) was heated at reflux in toluene (15 mL) for 30 min. The electrophile (1.9 mmol) and n-Bu₄NBr (1.2 mmol) were then added and stirring with heating continued until there was no change in the progress of the reaction as monitored by TLC. The reaction mixture was cooled, concentrated, and the products were purified by column chromatography.

Allyl 3,5-di-*O*-benzyl-β-D-arabinofuranoside (14) and 2-*O*-Allyl-3,5-di-*O*-benzyl-D-arabinofuranose (15). The crude mixture resulting from the reaction of 10 and allyl bromide was purified by chromatography (hexane/EtOAc, 5:1) to give 14 (39%) and 15 (24%) as clear oils. 14: R_f 0.5 (hexane/EtOAc, 2:1); $[\alpha]_D - 34.9^\circ$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.33–7.27 (m, 10 H), 5.88–5.81 (m, 1 H), 5.23 (ddd, 1 H, *J* = 17.2, 3.1, 1.6 Hz), 5.17 (ddd, 1 H, *J* = 10.4, 2.7, 1.3 Hz), 5.00 (d, 1 H, *J* = 4.7 Hz), 4.75 (d, 1 H, *J* = 11.9 Hz), 4.65 (d, 1 H, *J* = 12.3 Hz), 4.60–4.52 (m, 2 H), 4.26 (m, 2 H), 4.14 (dd, 1 H, *J* = 11.3, 5.6 Hz), 4.04–3.99 (m, 1 H), 3.86 (t, 1 H, *J* = 5.8 Hz), 3.53 (d, 2 H, *J* = 5.7 Hz), 2.62 (br d, 1 H, *J* = 7.9 Hz); ¹³C NMR (100 MHz CDCl₃, δ_C) 138.5 (2), 134.2, 129.0, 128.8. 128.2, 128.1, 118.1, 101.2, 85.2, 81.3, 78.5, 73.7, 72.5, 72.3, 69.1.

Anal. Calcd for C₂₂H₂₆O₅ (370.18): C, 71.33; H, 7.07. Found: C, 71.39; H, 7.10.

15: $R_f 0.4$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.35–7.25 (m, 10 H), 5.91–5.79 (m, 1 H), 5.34–5.16 (m, 3 H), 4.62–4.45 (m, 4 H), 4.11–3.89 (m, 5 H), 3.80 (d, 0.4 H, J = 6.0 Hz), 3.58–3.49 (m, 2 H), 3.32 (d, 0.6 H, J = 7.9 Hz); ¹³C NMR (100 MHz, CDCl₃, δ_C) 138.5, 138.3, 137.9, 137.7, 134.4, 134.3, 128.9, 128.8, 128.4, 128.3, 128.2 (2), 128.1, 118.3, 117.9, 101.5, 96.6, 86.8, 84.5, 83.1, 82.4, 82.2, 80.8, 74.0, 73.8, 72.5, 72.4, 71.7, 71.1, 70.9, 70.6. HRMS (ESI) Calcd for C₂₂H₂₆O₅ [M + Na]⁺ 393.1672. Found 393.1709.

Benzyl 3,5-di-*O*-benzyl-β-D-arabinofuranoside (16) and 2,3,5-Tri-*O*-benzyl-Darabinofuranose (17). The crude mixture resulting from the reaction of 10 and benzyl bromide was purified by chromatography (hexane/EtOAc, 4:1) to give 16 (39%) as a clear oil and $17^{[27]}$ (22%) as a white solid. 16: R_f 0.5 (hexane/EtOAc, 2:1); $[\alpha]_D - 40.2^\circ$ (*c* 0.02, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.35–7.19 (m, 15 H), 5.07 (d, 1 H, *J* = 4.7 Hz), 4.78 (d, 1 H, *J* = 8.0 Hz), 4.75 (d, 1 H, *J* = 8.3 Hz), 4.63– 4.48 (m, 4 H), 4.27 (t, 1 H, *J* = 5.4 Hz), 4.17 (dd, 1 H, *J* = 11.3, 5.6 Hz), 3.89 (t, 1 H, *J* = 5.9 Hz), 3.56 (d, 2 H, *J* = 5.7 Hz), 2.58 (br s, 1 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 138.0 (2), 137.1, 128.5, 128.4 (2), 128.2, 128.0, 127.8, 127.7 (2), 100.8, 84.7, 81.0, 78.1, 73.3, 72.0 (2), 69.7.

Anal. Calcd for C₂₆H₂₈O₅ (420.19): C, 74.26; H, 6.71. Found: C, 74.05; H, 6.66.

17: $R_f 0.3$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.34–7.24 (m, 15 H), 5.38 (d, 0.2 H, J = 7.7 Hz), 5.32 (dd, 0.8 H, J = 9.9, 4.3 Hz), 4.66–4.44 (m, 6 H), 4.17–4.14 (m, 0.8 H), 4.10–4.07 (m, 0.8 H), 4.02–3.97 (m, 1.4 H), 3.85 (d, 0.8 H, J = 10.0 Hz), 3.62–3.49 (m, 2 H), 3.23 (d, 0.2 H, J = 7.8 Hz); ¹³C NMR (100 MHz, CDCl₃, δ_C) 138.5, 138.3, 137.86, 137.9, 137.7, 129.0, 128.9, 128.8, 128.5, 128.4, 128.3 (2), 128.2 (2), 128.1, 101.6, 96.7, 86.9, 84.5, 83.1, 82.5, 82.3, 81.0, 77.8, 74.0, 73.8, 72.7, 72.5, 72.4, 72.2, 71.0, 70.6. HRMS (ESI) Calcd for C₂₆H₂₈O₅ [M + Na]⁺ 443.1834. Found 443.1829.

p-Methoxybenzyl 3,5-di-*O*-benzyl- β -D-arabinofuranoside (18) and 3,5-Di-*O*-benzyl-2-*O*-*p*-methoxybenzyl-D-arabinofuranose (19). The crude mixture resulting from the reaction of 10 and *p*-methoxybenzyl chloride was purified by chromatography (hexane/EtOAc, 3:1) to give 18 (36%) and 19 (36%) both as white solids. 18: R_f 0.3

(hexane/EtOAc, 2:1); mp 41–44°C; $[\alpha]_D - 25.5^{\circ}$ (*c* 0.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.33–7.25 (m, 10 H), 7.18 (d, 2 H, *J* = 8.4 Hz), 6.83 (d, 2 H, *J* = 8.4 Hz), 5.04 (d, 1 H, *J* = 4.6 Hz), 4.75 (d, 1 H, *J* = 11.9 Hz), 4.70 (d, 1 H, *J* = 11.2 Hz), 4.62–4.55 (m, 3 H), 4.42 (d, 1 H, *J* = 11.2 Hz), 4.27–4.23 (m, 1 H), 4.17 (dd, 1 H, *J* = 11.2, 5.6 Hz), 3.87 (t, 1 H, *J* = 5.9 Hz), 3.78 (s, 3 H), 3.56 (d, 2 H, *J* = 5.6 Hz), 2.62 (d, 1 H, *J* = 9.5 Hz); ¹³C NMR (100 MHz, CDCl₃, δ_C) 159.9, 138.5 (2), 130.4, 129.6, 128.9, 128.8, 128.2, 128.1 (2), 114.3, 101.0, 85.1, 81.3, 78.5, 73.8, 72.5, 72.3, 69.8, 55.7.

Anal. Calcd for $C_{27}H_{30}O_6$ (450.20): C, 71.98; H, 6.71. Found: C, 71.87; H, 6.68.

19: $R_f 0.2$ (hexane/EtOAc, 2:1); ¹H NMR (500 MHz, CDCl₃, δ_H) 7.40–7.30 (m, 10 H), 7.24 (d, 1 H, J = 6.9 Hz), 6.96–6.91 (m, 3 H), 5.40–5.35 (m, 1 H), 4.66–4.54 (m, 6 H), 4.52–4.42 (m, 1 H), 4.20–4.12 (m, 1 H), 4.06–3.95 (m, 1 H), 3.86 (s, 3 H), 3.66–3.52 (m, 2 H), 3.40 (br s, 1 H); ¹³C NMR (125 MHz, CDCl₃, δ_C) 160.0, 159.9, 159.7, 138.5, 138.3, 137.9, 137.8, 133.6, 130.2, 129.9, 129.8, 129.1, 128.9, 128.9, 128.8, 128.4, 128.3 (3), 128.2, 128.20, 128.1, 114.4 (2), 101.6, 96.7, 86.7, 84.1, 83.2, 82.4, 82.3, 80.9, 74.0, 73.78, 72.4 (2), 71.9, 71.0, 70.7, 65.4, 55.7 (2). HRMS (ESI) Calcd for C₂₇H₃₀O₆ [M + Na]⁺ 473.1940. Found 473.1941.

Allyl 3,5-di-*O*-benzoyl-β-D-arabinofuranoside (20) and 2-*O*-Allyl-3,5-di-*O*-benzoyl-D-arabinofuranose (21). The crude mixture resulting from the reaction of **11** and allyl bromide was purified by chromatography (hexane/EtOAc, 5:1) to give **20** (13%) and **21** (7%) as white solids. **20**: R_f 0.4 (hexane/EtOAc, 2:1); mp 60–64°C; $[\alpha]_D - 54.1^\circ$ (*c* 0.03, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.06–8.04 (m, 4 H), 7.61–7.37 (m, 2 H), 7.47–7.37 (m, 4 H), 5.94–5.84 (m, 1 H), 5.49 (t, 1 H, *J* = 6.0 Hz), 5.28 (ddd, 1 H, *J* = 17.2, 2.9, 1.4 Hz), 5.20 (dd, 1 H, *J* = 10.4 Hz), 5.15 (d, 1 H, *J* = 4.7 Hz), 4.67 (dd, 1 H, *J* = 11.7, 4.5 Hz), 4.55 (dd, 1 H, *J* = 11.7, 6.5 Hz), 4.50 (dd, 1 H, *J* = 6.5, 4.7 Hz), 4.43–4.39 (m, 1 H), 4.34–4.29 (m, 1 H), 4.12–4.07 (m, 1 H), 2.55 (br s, 1 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 166.5, 166.2, 133.5, 133.4, 133.0, 129.9 (2), 129.8, 129.2, 128.5, 128.4, 118.2, 100.5, 79.9, 79.3, 76.7, 69.0, 65.6.

Anal. Calcd for C₂₂H₂₂O₇ (398.1366): C, 66.32; H, 5.57. Found: C, 66.08; H, 5.70.

21: $R_f 0.3$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.09–8.04 (m, 4 H), 7.59–7.40 (m, 6 H), 5.94–5.87 (m, 1 H), 5.57–5.20 (m, 4 H), 4.73–4.55 (m, 3 H), 4.26–4.12 (m, 3 H), 3.90 (d, 0.3 H, J = 9.6 Hz), 2.77 (d, 0.7 H, J = 4.2 Hz); ¹³C NMR (100 MHz, CDCl₃, δ_C) 166.2, 166.1, 134.1 (2), 133.7, 133.9, 133.5, 130.3, 130.2, 129.0 (2), 128.8 (2), 118.3, 102.1, 88.7, 88.5, 87.7, 87.4, 78.6, 71.4, 65.5, 64.9.

Anal. Calcd for C₂₂H₂₂O₇ (398.1366): C, 66.32; H, 5.57. Found: C, 66.06; H, 5.70.

Benzyl 3,5-di-*O*-benzoyl-β-D-arabinofuranoside (22) and 3,5-di-*O*-benzoyl-2-*O*-benzyl-D-arabinofuranose (23). The crude mixture resulting from the reaction of 11 and benzyl bromide was purified by chromatography (hexane/EtOAc, 5:1) to give 22 (34%) and 23 (8%) both as white solids. 22: R_f 0.5 (hexane/EtOAc, 2:1); mp 85–87°C; $[\alpha]_D - 48.1^\circ$ (c 0.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.05–8.03 (m, 4 H), 7.59–7.29 (m, 11 H), 5.51 (t, 1 H, J = 6.1 Hz), 5.19 (d, 1 H, J = 4.7 Hz), 4.86 (d, 1

H, J = 11.7 Hz), 4.68 (dd, 1 H, J = 11.7, 4.4 Hz), 4.60 (d, 1 H, J = 11.7 Hz), 4.58 (dd, 1 H, J = 11.7, 6.6 Hz), 4.50 (ddd, 1 H, J = 10.1, 5.4, 3.6 Hz), 4.44–4.40 (m, 1 H), 2.94 (d, 1 H, J = 10.3 Hz); ¹³C NMR (100 MHz, CDCl₃, $\delta_{\rm C}$) 166.8, 166.6, 137.0, 133.9, 133.5, 130.3 (2), 129.7, 129.0, 128.9, 128.8, 128.7, 128.6, 100.9, 80.2, 79.8, 77.1, 70.5, 66.1.

Anal. Calcd for C₂₆H₂₄O₇ (448.1522): C, 69.63; H, 5.39. Found: C, 69.37; H, 5.53.

23: R_f 0.4 (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.06–8.03 (m, 4 H), 7.48–7.25 (m, 11 H), 5.60–5.46 (m, 2 H), 4.89–4.56 (m, 5 H), 4.38–4.34 (m, 0.3 H), 4.20 (br s, 0.4 H), 4.12 (dd, 0.3 H, J = 4.0, 2.7 Hz), 3.90 (br s, 0.3 H), 2.80 (br s, 0.7 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 166.8, 166.3, 137.6, 137.0, 134.1, 134.0, 133.4, 130.3, 130.2 (2), 129.7, 129.1, 129.0 (2), 128.9, 128.8, 128.7, 128.6, 128.4, 128.3, 102.1, 97.7, 87.6, 81.9, 81.5, 79.5, 78.5, 77.1, 73.3, 72.5, 65.5, 64.8. HRMS (ESI) Calcd for C₂₆H₂₄O₇ [M + Na]⁺ 471.1320. Found 471.1432.

p-Methoxybenzyl-3,5-di-*O*-benzoyl-β-D-arabinofuranoside (24) and 3,5-Di-*O*-benzoyl-2-*O*-*p*-methoxybenzyl-D-arabinofuranose (25). The crude mixture resulting from the reaction of **11** and *p*-methoxybenzyl chloride was purified by chromatography (hexane/EtOAc, 5:1) to give **24** (35%) and **25** (24%) both as white solids. **24**: R_f 0.3 (hexane/EtOAc, 2:1); mp 60–64°C; $[\alpha]_D - 42.2^\circ$ (*c* 0.03, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.11–8.09 (m, 4 H), 7.63 (t, 1 H, *J* = 7.4 Hz), 7.58 (t, 1 H, *J* = 7.4 Hz), 7.49 (t, 2 H, *J* = 7.8 Hz), 7.42 (t, 2 H, *J* = 7.8 Hz), 7.30 (d, 2 H, *J* = 8.6 Hz), 6.90 (d, 2 H, *J* = 8.5 Hz), 5.54 (t, 1 H, *J* = 6.0 Hz), 5.23 (d, 1 H, *J* = 4.7 Hz), 4.85 (d, 1 H, *J* = 11.3 Hz), 4.73 (dd, 1 H, *J* = 11.7, 4.3 Hz), 4.64 (dd, 1 H, *J* = 11.7, 6.7 Hz), 4.59 (d, 1 H, *J* = 11.3 Hz), 4.54 (dd, 1 H, *J* = 6.4, 4.8 Hz), 4.49–4.45 (m, 1 H), 3.84 (s, 3 H), 2.67 (br s, 1 H); ¹³C NMR (125 MHz, CDCl₃, δ_C) 166.8, 166.6, 160.0, 133.9, 133.4 (2), 130.3, 130.2, 129.7, 129.1, 128.9, 128.8, 114.4, 100.6, 80.3, 79.9, 77.1, 70.2, 66.1, 55.7. HRMS (ESI) Calcd for C₂₇H₂₆O₈ [M + Na]⁺ 501.1525. Found 501.1521.

25: R_f 0.2 (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.06–8.03 (m, 4 H), 7.47–7.26 (m, 8 H), 6.86–6.83 (m, 2 H), 5.56–5.44 (m, 2 H), 4.81–4.55 (m, 4.5 H), 4.37–4.33 (m, 0.5 H), 4.17 (s, 0.6 H), 4.09 (dd, 0.5 H, J = 4.0, 2.6 Hz), 3.96 (br d, 0.4 H J = 8.2 Hz), 3.77 (s, 3 H), 2.92 (br s, 0.5 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 166.3, 165.9, 159.4, 133.6, 133.5, 133.1, 133.0, 129.9 (2), 129.8, 129.6, 129.3 (2), 129.2, 128.7, 128.6 (2), 128.5, 128.4, 128.3, 114.1, 113.9, 101.8, 97.3, 86.9, 81.4, 80.7, 79.12, 78.17, 76.73, 72.62, 71.79, 65.1, 64.5, 55.3. HRMS (ESI) Calcd for C₂₇H₂₆O₈ [M + Na]⁺ 501.1525. Found 501.1520.

Allyl 3,5-*O*-(tetraisopropylsiloxane-1,3-diyl)-β-D-arabinofuranoside (26) and 3,5-*O*-(tetraisopropylsiloxane-1,3-diyl)-2-*O*-benzyl-D-arabinofuranose (27). The crude reaction mixture resulting from the reaction of 12 and allyl bromide was purified by chromatography (hexane/EtOAc, 6:1) to give 26 (23%) and 27 (36%) as clear oils. 26: R_f 0.8 (hexane/EtOAc, 2:1); $[\alpha]_D - 58.1^\circ$ (*c* 0.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 5.94–5.84 (m, 1 H), 5.27 (ddd, 1 H, J = 17.2, 3.1, 1.5 Hz), 5.20 (ddd, 1 H, J = 10.4, 2.7, 1.3 Hz), 4.90 (d, 1 H, J = 4.7 Hz), 4.26–4.20 (m, 2 H), 4.16–4.10 (m, 1 H), 4.03–3.99 (m, 1 H), 3.95 (dd, 1 H, J = 10.7, 3.2 Hz), 3.88–3.83 (m, 1 H), 3.78 (dd, 1 H, J = 10.7, 8.8 Hz), 2.30 (d, 1 H, J = 10.3 Hz), 1.11–0.94 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, $\delta_{\rm C}$) 134.3, 117.9, 99.9, 82.6, 80.0, 79.1, 68.8, 66.4, 18.0, 17.9, 17.80, 17.5, 17.4 (2), 13.9, 13.7, 13.72, 13.3, 12.9.

Anal. Calcd for C₂₀H₄₀O₆Si₂ (432.24): C, 55.52; H, 9.32. Found: C, 55.67; H, 9.32.

27: $R_f 0.7$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, $\delta_{\rm H}$) 5.96–5.85 (m, 1 H), 5.36–5.32 (m, 1 H), 5.32–5.28 (m, 1 H), 5.25–5.18 (m, 1 H), 4.34 (t, 0.5 H, J = 6.2 Hz), 4.26 (dd, 0.5 H, J = 6.9, 5.0 Hz), 4.19–4.05 (m, 2.5 H), 4.03–3.98 (m, 1 H), 3.93–3.84 (m, 2 H), 3.81–3.77 (m, 0.5 H), 3.32 (d, 0.5, J = 6.2 Hz), 2.82 (d, 0.5 H, J = 5.2 Hz), 1.10–1.02 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, $\delta_{\rm C}$) 134.3, 134.0, 117.9, 117.1, 100.5, 94.3, 89.5, 84.7, 81.8, 81.2, 76.7, 76.5, 72.0, 71.4, 65.5, 62.4, 17.4 (5), 17.1 (4), 17.0, 13.5 (2), 13.1, 13.0, 12.9, 12.6, 12.5. HRMS (ESI) Cald for $C_{20}H_{40}O_6Si_2$ [M + Na]⁺ 455.2256. Found 455.2236.

Benzyl 3,5-*O*-(tetraisopropylsiloxane-1,3-diyl)-β-D-arabinofuranoside (28) and 3,5-*O*-(Tetraisopropylsiloxane-1,3-diyl)-2-*O*-benzyl-D-arabinofuranose (29). The crude mixture resulting from the reaction of 12 and benzyl bromide was purified by chromatography (hexane/EtOAc, 6:1) to give $28^{[43]}$ (36%) as a white solid and 29 (37%) as a clear oil. 28: R_f 0.8 (hexane/EtOAc, 2:1); $[\alpha]_D - 113.2^\circ$ (*c* 0.02, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.38–7.29 (m, 5 H), 4.97 (d, 1 H, *J* = 4.7 Hz), 4.77 (d, 1 H, *J* = 11.5 Hz), 4.50 (d, 1 H, *J* = 11.5 Hz), 4.26 (dd, 1 H, *J* = 7.4, 6.0 Hz), 4.17–4.12 (m, 1 H), 3.97 (dd, 1 H, 10.5 Hz), 1.09–0.94 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 137.1, 128.5, 128.2, 128.0, 99.6, 82.2, 79.6, 78.7, 69.6, 66.0, 17.6, 17.5, 17.4 (2), 17.2, 17.0 (3), 13.5, 13.3, 12.9, 12.5.

Anal. Calcd for C₂₄H₄₂O₆Si₂ (482.25): C, 59.71; H, 8.77. Found: C, 59.95; H, 8.77.

29: $R_f 0.7$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.36–7.30 (m, 5 H), 5.34 (dd, 0.7 H, J = 6.6, 2.5 Hz), 5.23 (dd, 0.3 H, J = 7.7, 5.5 Hz), 4.75–4.60 (m, 2 H), 4.43 (t, 0.3 H, J = 7.6 Hz), 4.34 (dd, 0.7 H, J = 8.5, 6.0 Hz), 4.15–4.09 (m, 0.8 H), 4.06 (d, 0.3 H), 4.03–3.98 (m, 1.7 H), 3.92–3.86 (m, 1 H), 3.86–3.80 (m, 0.2 H), 3.33 (d, 0.3 H, J = 6.2 Hz), 2.83 (d, 0.7 H, J = 5.3 Hz). 1.10–0.91 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 138.2, 137.7, 128.9, 128.8, 128.5, 128.3, 128.1 (2), 100.9, 94.7, 89.9, 85.1, 82.5, 81.8, 77.8, 77.1, 73.2, 72.7, 66.0, 62.9, 17.9, 17.8 (3), 17.5 (4), 17.4 (2), 13.9 (2), 13.7, 13.5, 13.3, 13.0. HRMS (ESI) Calcd for C₂₄H₄₂O₆Si₂ [M + Na]⁺ 505.2412. Found 505.2432.

p-Methoxybenzyl 3,5-*O*-(tetraisopropylsiloxane-1,3-diyl)-β-D-arabinofuranoside (30) and 3,5-*O*-(Tetraisopropylsiloxane-1,3-diyl)-2-*O*-*p*-methoxybenzyl-D-arabinofuranose (31). The crude mixture resulting from the reaction of 12 and *p*-methoxybenzyl chloride was purified by chromatography (hexane/EtOAc, 6:1) to give 30 (45%) and 31 (36%) as clear oils. 30: R_f 0.7 (hexane/EtOAc, 2:1); $[\alpha]_D - 80.9^\circ$ (*c* 0.09, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.26–7.23 (m, 2 H), 6.89–6.87 (m, 2 H), 4.95 (d, 1 H, *J* = 4.7 Hz), 4.70 (d, 1 H, *J* = 11.2 Hz), 4.43 (d, 1 H, *J* = 11.2 Hz), 4.23 (dd, 1 H, *J* = 7.3, 5.9 Hz), 4.15–4.09 (m, 1 H), 3.97 (dd, 1 H, *J* = 10.7, 3.2 Hz),

3.89-3.85 (m, 1 H), 3.81 (s, 3 H), 3.80 (dd, 1 H, J = 10.7, 8.8 Hz), 2.28 (d, 1 H, J = 10.4 Hz), 1.10–0.95 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, $\delta_{\rm C}$) 159.9, 130.3, 129.6, 114.3, 99.7, 82.6, 80.1, 79.1, 69.7, 66.5, 55.7, 18.0, 17.8 (2), 17.4 (3), 13.9, 13.7, 13.3, 12.9.

Anal. Calcd for C₂₅H₄₄O₇Si₂ (512.26): C, 58.56; H, 8.65. Found: C, 58.38; H, 8.59.

31: $R_f 0.5$ (hexane/EtOAc, 2:1); ¹H NMR (400 MHz, CDCl₃, δ_H) 7.29–7.25 (m, 2 H), 6.89–6.86 (m, 2 H), 5.30 (dd, 0.6 H, J = 6.6, 2.4 Hz), 5.16 (t, 0.4 H, J = 3.8 Hz), 4.65–4.55 (m, 2 H), 4.37 (t, 0.4 H, J = 7.6 Hz), 4.29 (dd, 0.6 H, J = 8.4, 6.0 Hz), 4.09-4.03 (m, 1 H), 4.01-3.97 (m, 1 H), 3.95-3.93 (m, 1 H), 3.88-3.83 (m, 1 H), 3.81 (s, 3 H), 3.33 (d, 0.4 H, J = 6.2 Hz), 2.83 (d, 0.6 H, J = 5.4 Hz), 1.08–0.94 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 160.0, 159.7, 130.2, 130.1, 129.8, 114.3, 114.2, 101.0, 94.8, 89.6, 84.9, 82.7, 81.8, 77.9, 77.2, 77.1, 73.0, 72.5, 65.9, 63.1, 55.7 (2), 18.0, 17.9, 17.8 (3), 17.6, 17.5 (2) 17.4, 13.9, 13.7 (2), 13.4, 13.0, 12.9. HRMS (ESI) Calcd for $C_{25}H_{44}O_7Si_2$ [M + Na]⁺ 535.2518. Found 535.2494.

Methyl 2.3-anhydro-5-O-toluenesulfonyl-q-D-lyxofuranoside (34). Alcohol 32 (0.90 g, 6.2 mmol) and p-TsCl (2.41 g, 12.7 mmol) were dissolved in pyridine and stirred overnight at room temperature. The mixture was diluted with CH₂Cl₂, washed with water, and the organic layer was dried (MgSO₄) and concentrated. Column chromatography (hexane/EtOAc, 2:1) of the residue gave 34 (1.55 g, 83%) as a white solid: R_f 0.4 (hexane/EtOAc, 2:1); mp 74–76°C.^[44] ¹H NMR (400 MHz, CDCl₃, δ_H) 7.81 (d, 2 H, J = 8.3 Hz), 7.36 (d, 2 H, J = 8.0 Hz), 4.88 (s, 1 H), 4.21 (t, 1 H, J = 6.3Hz), 4.14–4.12 (m, 2 H), 3.72 (d, 1 H, J = 2.8 Hz), 3.64 (d, 1 H, J = 2.8 Hz), 3.37 (s, 3 H), 2.45 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 145.1, 132.6, 130.0, 128.0, 102.3, 73.7, 67.5, 56.5, 55.7, 53.7, 21.7.

Methyl 2,3-anhydro-5-deoxy-5-iodo- α -D-lyxofuranoside (35). Tosylate 34 (1.18 g, 3.9 mmol) and *n*-Bu₄I (3.03 g, 8.2 mmol) were dissolved in toluene (20 mL) and heated at reflux for 7 h. The mixture was concentrated and the residue was dissolved in CH₂Cl₂, washed with water and a saturated aqueous solution of NaHSO₃. The organic layer was dried $(MgSO_4)$ and concentrated. Column chromatography (hexane/EtOAc, 4:1) of the residue gave 35 (0.86 g, 86%) as a white solid: R_f 0.6 (hexane/EtOAc, 2:1). $[\alpha]_D$ + 68.7° (c 0.2, CHCl₃); mp 74–76°C; ¹H NMR (400 MHz, $CDCl_3$, δ_H) 5.05 (s, 1 H), 4.28 (t, 1 H, J = 7.2 Hz), 3.90 (d, 1 H, J = 2.8 Hz), 3.68 (d, 1 H, J = 2.8 Hz), 3.42 (s, 3 H), 3.24 (s, 1 H), 3.23 (d, 1 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 102.3, 76.7, 56.3, 55.0, 54.4, 0.6. HRMS (ESI) Calcd for C₆H₉IO₃ $[M + Na]^+$ 278.9489. Found 278.9481.

Methyl 2,3-anhydro-5-O-formyl-a-D-lyxofuranoside (37). A mixture of 12 (0.15 g, 0.38 mmol) and dibutyltin oxide (0.10 g, 0.41 mmol) in toluene (12 mL) was heated at reflux for 20 min. Triflate 33 (0.31 g, 1.12 mmol) was added and stirring continued at reflux for 6 h before DMF (2 mL) was added. After 1 h, a new product was detected by TLC, and the reaction mixture was then cooled and concentrated. The residue was dissolved in CH₂Cl₂, washed with water, and the organic layer was dried

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016 (Na₂SO₄), and concentrated. The residue was purified by chromatography (hexane/EtOAc, 6:1) to give **38** (77 mg, 39%) as a white solid: R_f 0.4 (hexane/EtOAc, 2:1); $[\alpha]_D$ + 13.8° (*c* 0.02, CHCl₃); ¹H NMR (400 MHz, CDCl₃, δ_H) 8.10 (d, 1 H, J = 0.7 Hz,), 4.98 (s, 1 H), 4.37 (d, 2 H, J = 6.2 Hz), 4.23 (t, 1 H, J = 5.9 Hz), 3.74 (dd, 1 H, J = 2.8, 0.7 Hz), 3.68 (d, 1 H, J = 2.8 Hz), 3.43 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃, δ_C) 160.9, 102.8, 74.2, 62.5, 56.6, 56.2, 54.2. HRMS (ESI) Calcd for C₇H₁₀O₅ [M + Na]⁺ 197.0420. Found 197.0424.

Computational studies on 43. Because many force fields for stochastic searching do not effectively handle tin as an element, a surrogate approach using silicon as a mimic was employed. Thus, the SPMC search protocol available in MacroModel Version 6.5 was used to generate an initial family of 10,000 conformers of 44. Each conformer was then minimized in the gas phase using AM1,^[40] which generated 113 unique conformers, all of which were within 32 kcal/mol of the global minimum. Two families of structures were present. One family (four conformers) was clustered at the global minimum and the other (6 conformers) ~ 1.1 kcal/mol above the global minimum. Within each family of structures, the furanose ring adopted the same conformation, and the geometrical differences were only in the orientations about the bonds in the siloxane moeity. The two lowest energy conformers in each family were subsequently optimized at the B3LYP level of theory after replacement of the surrogate silicon atom with tin. The basis set used for the full geometry optimization was 3-21G* for silicon, oxygen, carbon, and hydrogen and the basis set for tin was the SDD effective core potential.^[45] Single-point energy calculations were subsequently performed at the B3LYP level of theory^[46-48] using a 6-31G* basis set for silicon, oxygen, carbon, and hydrogen and the SDD effective core potential for tin. All DFT calculations utilized 6 Cartesian d functions and were performed using Gaussian 98.^[49]

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REFERENCES

- Brennan, P.J.; Nikaido, H. The envelope of mycobacteria. Ann. Rev. Biochem. 1995, 64, 29.
- Lowary, T.L. Mycobacterial cell wall components. In *Glycoscience: Chemistry and Chemical Biology*; Fraser-Reid, B., Tatsuta, K., Thiem, J., Eds.; Springer-Verlag: Berlin, 2001; 2005–2080.
- 3. Lowary, T.L. Recent progress towards the identification of inhibitors of mycobacterial cell wall polysaccharide biosynthesis. Mini-Rev. Med. Chem. **2003**, *3*, 694.

Downloaded At: 07:00 23 January 2011

- 4. Crick, D.C.; Mahapatra, S.; Brennan, P.J. Biosynthesis of the arabinogalactanpeptidoglycan complex of *Mycobacterium tuberculosis*. Glycobiology **2001**, *11*, 107R.
- 5. Mereyala, H.B.; Hotha, S.; Gurjar, M.K. Synthesis of pentaarabinofuranosyl structure motif A of *Mycobacterium tuberculosis*. J. Chem. Soc., Chem. Commun. **1998**, 685.
- 6. Désiré, J.; Prandi, J. Synthesis of methyl β-D-arabinofuranoside 5-[1-D (and L)-*myo*inositol 1-phosphate], the capping motif of the lipoarabinomannan of *Mycobacterium smegmatis*. Carbohydr. Res. **1999**, *317*, 110.
- 7. Sanchez, S.; Bamhaoud, T.; Prandi, J. Elaboration of monoarabinofuranosidic building blocks. Eur. J. Org. Chem. **2002**, *22*, 3864.
- Subramaniam, V.; Lowary, T.L. Synthesis of oligosaccharide fragments of mannosylated lipoarabinomannan from *Mycobacterium tuberculosis*. Tetrahedron 1999, 55, 5965.
- Yin, H.; D'Souza, F.W.; Lowary, T.L. Arabinofuranosides from mycobacteria: synthesis of a highly branched hexasaccharide and related fragments containing βarabinofuranosyl residues. J. Org. Chem. 2002, 67, 892.
- 10. Gadikota, R.R.; Callam, C.S.; Wagner, T.; Del Fraino, B.; Lowary, T.L. 2,3-Anhydrosugars in glycoside bond synthesis. Highly stereocontrolled syntheses of oligosaccharides containing α and β -arabinofuranosyl linkages. J. Am. Chem. Soc. **2003**, *125*, 4155.
- Ayers, J.D.; Lowary, T.L.; Morehouse, C.B.; Besra, G.S. Synthetic arabinofuranosyl oligosaccharides as mycobacterial arabinosyltransferase substrates. Bioorg. Med. Chem. Lett. **1998**, *8*, 437.
- Kaur, D.; Lowary, T.L.; Vissa, V.D.; Crick, D.C.; Brennan, P.J. Characterization of the epitope of anti-lipoarabinomannan antibodies as the terminal hexaarabinofuranosyl motif of mycobacterial arabinans. Microbiology UK 2002, *148*, 3059.
- Pathak, A.K.; Pathak, V.; Suling, W.J.; Gurcha, S.S.; Morehouse, C.B.; Besra, G.S.; Maddry, J.A.; Reynolds, R.C. Studies on *n*-octyl-5-(α-D-arabinofuranosyl)-β-Dgalactofuranosides for mycobacterial glycosyltransferase activity. Bioorg. Med. Chem. **2002**, *10*, 923.
- 14. Pathak, A.K.; Pathak, V.; Maddry, J.A.; Suling, W.J.; Gurcha, S.S.; Besra, G.S.; Reynolds, R.C. Studies on α - $(1 \rightarrow 5)$ linked octyl arabinofuranosyl disaccharides for mycobacterial arabinosyltransferase activity. Bioorg. Med. Chem. **2001**, *9*, 3145.
- 15. Pathak, A.K.; Pathak, V.; Seitz, L.; Maddry, J.A.; Gurcha, S.S.; Besra, G.S.; Suling, W.J.; Reynolds, R.C. Studies on β - $(1 \rightarrow 5)$ and β - $(1 \rightarrow 6)$ linked octyl Gal*f* disaccharides as substrates for mycobacterial galactosyltransferase activity. Bioorg. Med. Chem. **2001**, *9*, 3129.
- Gridley, J.J.; Osborn, H.M.I. Recent advances in the construction of β-D-mannose and β-D-mannosamine linkages. J. Chem. Soc., Perkin Trans. 1 2000, 1471.
- 17. Pozsgay, V. Stereoselective synthesis of β-D-mannosides. In *Carbohydrates in Chemistry and Biology*; Ernst, B., Hart, G.W., Sinaÿ, P., Eds.; Wiley-VCH: Weinheim, 2000.
- Oscarson, S.; Sehgelmeble, F.W. Chemical syntheses of inulin and levan structures. J. Org. Chem. 2002, 67, 8457.



- Oscarson, S.; Sehgelmeble, F.W. A novel β-directing fructofuranosyl donor concept. Stereospecific synthesis of sucrose. J. Am. Chem. Soc. 2000, 122, 8869.
- Krog-Jensen, C.; Oscarson, S. Stereospecific synthesis of β-D-fructofuranosides using thioglycoside donors and internal aglycon delivery. J. Org. Chem. 1998, 63, 1780.
- Hodosi, G.; Kovác, P. A fundamentally new, simple, stereospecific synthesis of oligosaccharides containing the β-mannopyranosyl and β-rhamnopyranosyl linkage. J. Am. Chem. Soc. 1997, *119*, 2335.
- 22. Hodosi, G.; Kovác, P. Glycosylation via locked anomeric configuration: stereospecific synthesis of oligosaccharides containing the β -D-mannopyranosyl and β -L-rhamnopyranosyl linkage. Carbohydr. Res. **1998**, *308*, 63.
- 23. Srivastava, V.K.; Schuerch, C. Synthesis of β -D-mannopyrabnosides and regioselective *O*-alkylation of dibutylstannylene complexes. Tetrahedron Lett. **1979**, 3269.
- Dessinges, A.; Olesker, A.; Lukacs, G.; Thang, T.T. Carbohydrate synthesis for nuclear medicine—a new, rapid, and stereospecific route to 2-deoxy-2-fluoro-Dglucose. Carbohydr. Res 1984, 126, C6.
- 25. Su, T.-S.; Klein, R.A.; Fox, J.J. Improved synthesis of α -D-ribofuranosides via stereoselective alkylation of a dibutylstannylene derivative for ready access to the 2-substituted 2-deoxyarabinofuranosides. J. Org. Chem. **1982**, *47*, 1506.
- Ning, J.; Kong, F. Synthesis and glycosidic coupling reaction of substituted 2,6dioxabicyclo[3.1.0]hexanes: 1,2-anhydro-3,5-di-*O*-benzyl-α-D-ribofuranose. Carbohydr. Res. **1997**, *300*, 355.
- Reist, E.J.; Hart, P.A.; Goodman, L.; Baker, B.R. Potential anticancer agents. XXIII. The anomeric ethyl 1-thio-D-arabinofuranosides. J. Am. Chem. Soc. 1959, 81, 5176.
- D'Souza, F.W.; Ayers, J.D.; McCarren, P.R.; Lowary, T.L. Arabinofuranosyl oligosaccharides from mycobacteria: synthesis and effect of glycosylation on ring conformation and hydroxymethyl group rotamer populations. J. Am. Chem. Soc. 2000, 122, 1251.
- Mizutani, K.; Kasai, R.; Nakamura, M.; Tanaka, O.; Matsuura, H. NMR spectral study of α- and β-L-arabinofuranosides. Carbohydr. Res. 1989, 185, 27.
- 30. Gent, P.A.; Gigg, R. The allyl ether as a protecting group in carbohydrate chemistry: part VIII. Synthesis of benzyl and allyl ethers of D-glucopyranose. Carbohydr. Res. **1976**, *49*, 325.
- 31. Grindley, T.B. Applications of tin-containing intermediates to carbohydrate chemistry. Adv. Carbohydr. Chem. Biochem. **1998**, *53*, 17.
- 32. David, S.; Hanessian, S. Regioselective manipulation of hydroxyl groups via organotin derivatives. Tetrahedron **1985**, *41*, 643.
- Wagner, D.; Verheyden, J.P.H.; Moffatt, J.G. Preparation and synthetic utility of some organotin derivatives of nucleosides. J. Org. Chem. 1974, 39, 24.
- 34. Grouiller, A.; Buet, V.; Uteza, V.; Descotes, G. Novel *p*-toluenesulfonylation and thiocarbonylation of unprotected thymine nucleosides. Synlett **1993**, 221.
- Gough, G.R.; Miller, T.J.; Mantick, N.A. *p*-Nitrobenzyloxymethyl: a new fluorideremovable protecting group for ribonucleoside 2'-hydroxyls. Tetrahedron Lett. 1996, 37, 981.

980

- 36. Labanowski, J.W.: Andzelm, J. Density Functional Methods in Chemistry; Springer: New York, 1991.
- 37. Parr, R.G.; Yang, W. Density Functional Theory in Atoms and Molecules; Oxford University Press: New York, 1989.
- Mohamadi, F.; Richards, N.G.J.; Guida, W.C.; Liskamp, R.; Lipton, M.; Caulfield, 38. C.; Chang, G.; Hendrickson, T.; Still, W.C. Macromodel-an integrated software system for molecular modeling organic and bioorganic molecules using molecular mechanics. J. Comput. Chem. 1990, 11, 440.
- 39. Goodman, J.; Still, W.C. An unbounded systematic search of conformational space. J. Comput. Chem. 1991, 12, 1110.
- 40. Hehre, W.J.; Radom, L.; Schleyer, P.v.R.; Pople, J.A. Ab initio Molecular Orbital Theory; John Wiley & Sons: New York, 1986.
- Altona, C.; Sundaralingam, M. Conformational analysis of the sugar ring in 41. nucleosides and nucleotides. A new description using the concept of pseudorotation. J. Am. Chem. Soc. 1972, 94, 8205.
- 42. Martin, M.G.; Ganem, B.; Rasmussen, J.R. An improved synthesis of methyl 2,3anhydro-a-D-lyxofuranoside. Carbohydr. Res. 1983, 123, 332.
- 43. Kakinuma, H.; Yuasa, H.; Hashimoto, H. Glycosyltransfer mechanism of α glucosyltransferase from Protaminobacter rubrum. Carbohydr. Res. 1998, 312, 103.
- 44. Unger, F.M.; Christian, R.; Waldstaetten, P. Regiospecific syntheses of azidodeoxy and diazidodideoxy derivatives of methyl α -D-arabinofuranoside. Carbohydr. Res. **1979**, *69*, *71*.
- Andrae, D.; Haeussermann, U.; Dolg, M.; Stoll, H.; Preuss, H. Energy-adjusted ab 45. initio pseudopotentials for the second and third row transition elements. Theor. Chim. Acta 1990, 77, 123.
- 46. Becke, A.D. Density functional exchange energy approximation with correct asymptotic behavior. Phys. Rev., A 1988, 38, 3098.
- Becke, A.D. Density functional thermochemistry. 3. The role of exact exchange. J. 47. Chem. Phys. 1993, 98, 5648.
- 48. Lee, C.; Yang, W.; Parr, R.G. Development of the Colle-Salvetti correlation energy formula into a functional of the electron density. Phys. Rev., B 1988, 37, 785.
- 49. Frisch, M.J.; Trucks, G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Zakrzewski, V.G.; Montgomery, J.A., Jr.; Stratmann, R.E.; Burant, J.C.; Dapprich, S.; Millam, J.M.; Daniels, A.D.; Kudin, K.N.; Strain, M.C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G.A.; Ayala, P.Y.; Cui, Q.; Morokuma, K.; Malick, D.K.; Rabuck, A.D.; Raghavachari, K.; Foresman, J.B.; Cioslowski, J.; Ortiz, J.V.; Stefanov, B.B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R.L.; Fox, D.J.; Keith, T.; Al-Laham, M.A.; Peng, C.Y.; Nanayakkara, A.; Gonzalez, C.; Challacombe, M.; Gill, P.M.W.; Johnson, B.; Chen, W.; Wong, M.W.; Andres, J.L.; Gonzalez, C.; Head-Gordon, M.; Replogle, E.S.; Pople, J.A. Gaussian 98, Revision A.9; Gaussian, Inc.: Pittsburgh, PA, 1998.

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